A structured approach ... Part III Biological applications

David Gilbert

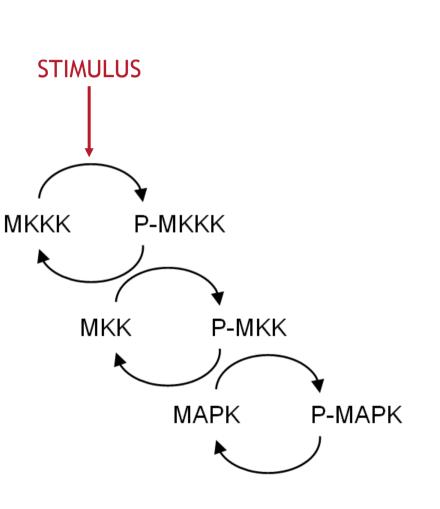
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Biological Applications

MAPK Pathway

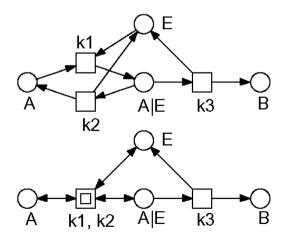
- Responds to wide range of stimuli: cytokines, growth factors, neurotransmitters, cellular stress and cell adherence,...
- Pivotal role in many key cellular processes:
 - growth control in all its variations,
 - cell differentiation and survival
 - cellular adaptation to chemical and physical stress.
- Deregulated in various diseases: cancer; immunological, inflammatory and degenerative syndromes,
- Represents an important drug target.

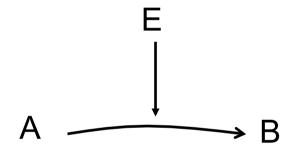


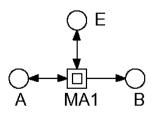
MA1: Mass action for enzymatic reaction - phosphorylation

$$E + A \xrightarrow[k_2]{k_1} E \mid A \xrightarrow[k_3]{k_3} E + B$$

- A: substrate
- B: product (phosphorylated A)
- E: enzyme (kinase)
- E|A substrate-enzyme complex







Differential equations

Enzymatic reaction

$$A + E \xrightarrow[k_2]{k_1} A \mid E \xrightarrow{k_3} B + E$$

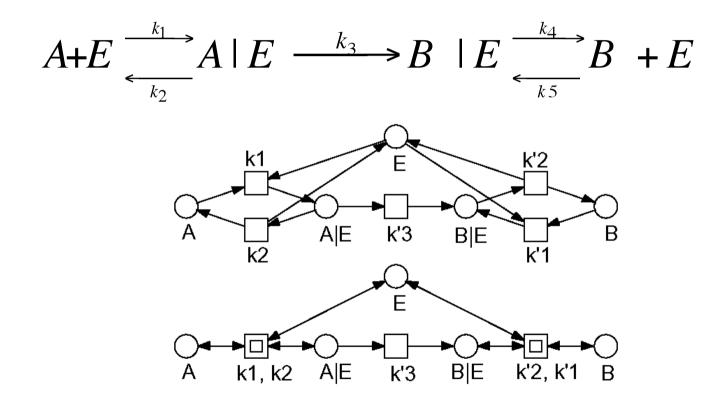
$$\frac{d[A]}{dt} = -k_1 \times [A] \times [E] + k_2 \times [A \mid E]$$

$$\frac{d[A \mid E]}{dt} = +k_1 \times [A] \times [E] - k_2 \times [A \mid E] - k_3 \times [A \mid E]$$

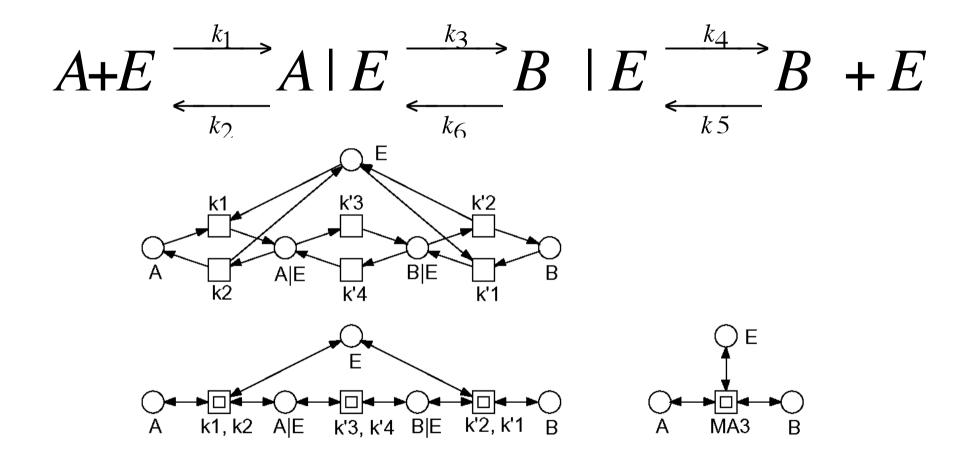
$$\frac{d[B]}{dt} = +k_3 \times [A \mid E] + k_3 \times [A \mid E]$$

$$\frac{d[E]}{dt} = -k_1 \times [A] \times [E] + k_2 \times [A \mid E] + k_3 \times [A \mid E]$$

MA2 model



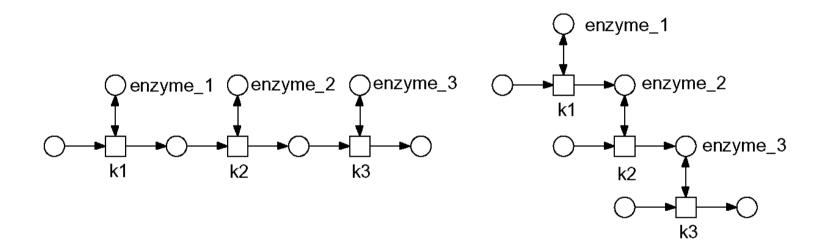
MA3 model



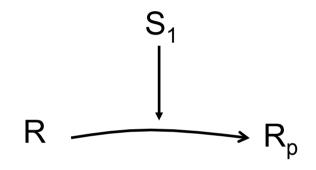
Multiple substrates

$$A_1 + A_2 + E \xrightarrow[k_{2A_1}]{k_{1A_1}} A_1 | E + A_2 \xrightarrow[k_{2A_2}]{k_{1A_2}} A_1 | A_2 | E \xrightarrow[k_{2A_2}]{k_{2A_2}} A_1 | A_2 | E \xrightarrow[k_{2A_2}]{k_{2A_2}} B_1 + B_2 + E$$

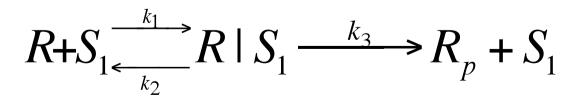
Metabolic pathways vs Signalling Pathways (Petri Nets)



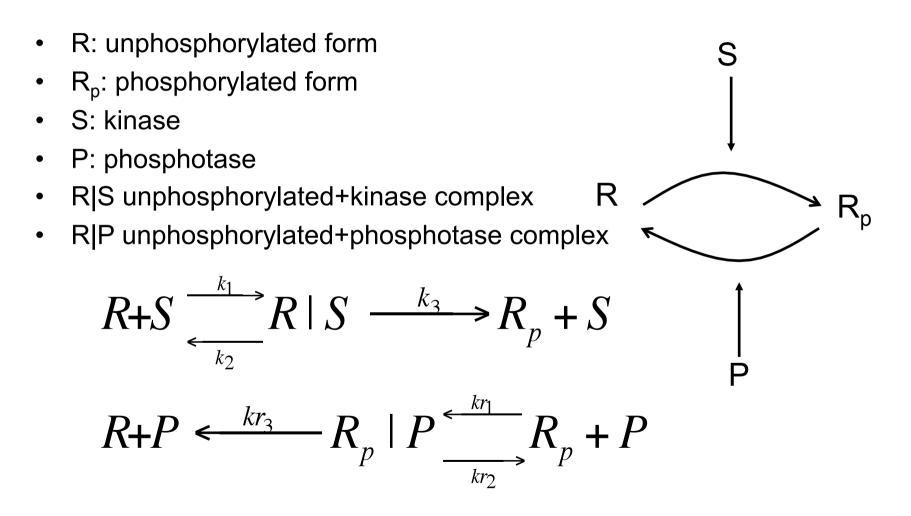
Mass action for enzymatic reaction - phosphorylation



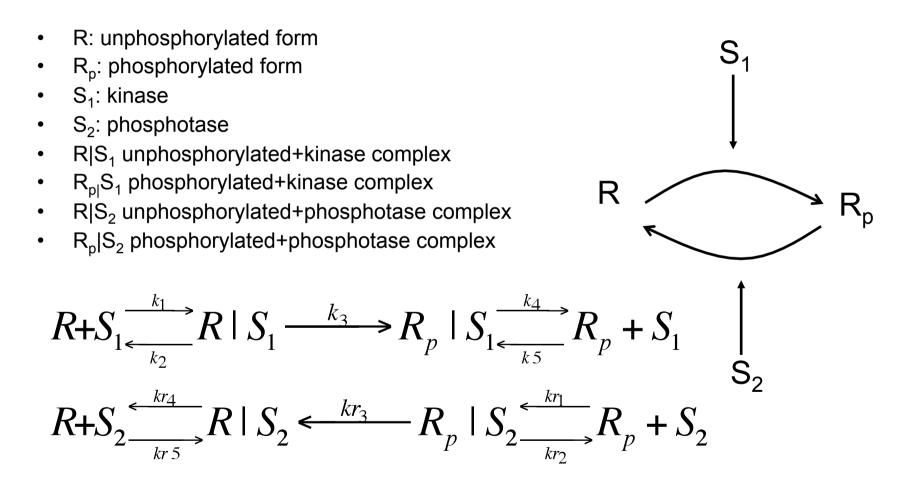
- R: substrate,
- R_p: product (phosphorylated R)
- S₁: enzyme (kinase)
- R|S₁ substrate-enzyme complex



Phosphorylation - dephosphorylation step Mass action model 1



Phosphorylation - dephosphorylation loop Mass action model 2



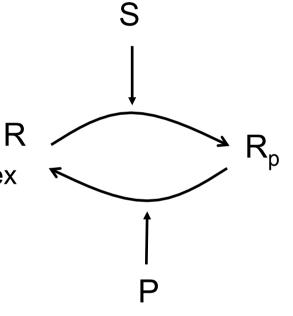
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Phosphorylation - dephosphorylation step Mass action (all singing/dancing)

- R: unphosphorylated form
- R_p: phosphorylated form
- S: kinase •
- P: phosphotase •
- RIS unphosphorylated+kinase complex •
- RIP unphosphorylated+phosphotase complex

$$R+S \xrightarrow[k_{1}]{k_{1}} R \mid S \xrightarrow[k_{6}]{k_{3}} R_{p} \mid S \xrightarrow[k_{5}]{k_{4}} R_{p} + S$$

$$R+P \xrightarrow[k_{7}]{k_{7}} R \mid P \xrightarrow[k_{7}]{k_{7}} R_{p} \mid P \xrightarrow[k_{7}]{k_{7}} R_{p} + R$$



kr5



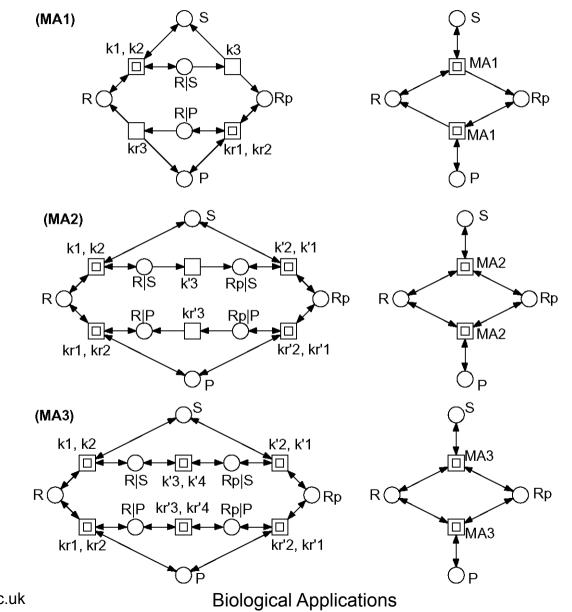
$$V = k_3 \times [S] \times \frac{[R]}{(K_{m1} + [R])} - k_3' \times \frac{[R_p]}{(K_{m2} + [R_p])}$$

- Assumptions:
 - 1. No product reverts to initial substrate
 - 2. MM Equation holds at initial stage of reaction before concentration of product is appreciable
 - 3. [Enzyme] << [Substrate]
- K_m is [Substrate] at which the reaction rate is half its maximum value
- $dR_p/dt == reaction rate V$
- $k_3 \ge S == V_{max}$ for the forward reaction
- $k_3' == V_{max}$ for the reverse reaction (Phosphotase is ignored)
- $K_{m1} == (k_2 + k_3)/k_1$ (k's from mass-action 1)

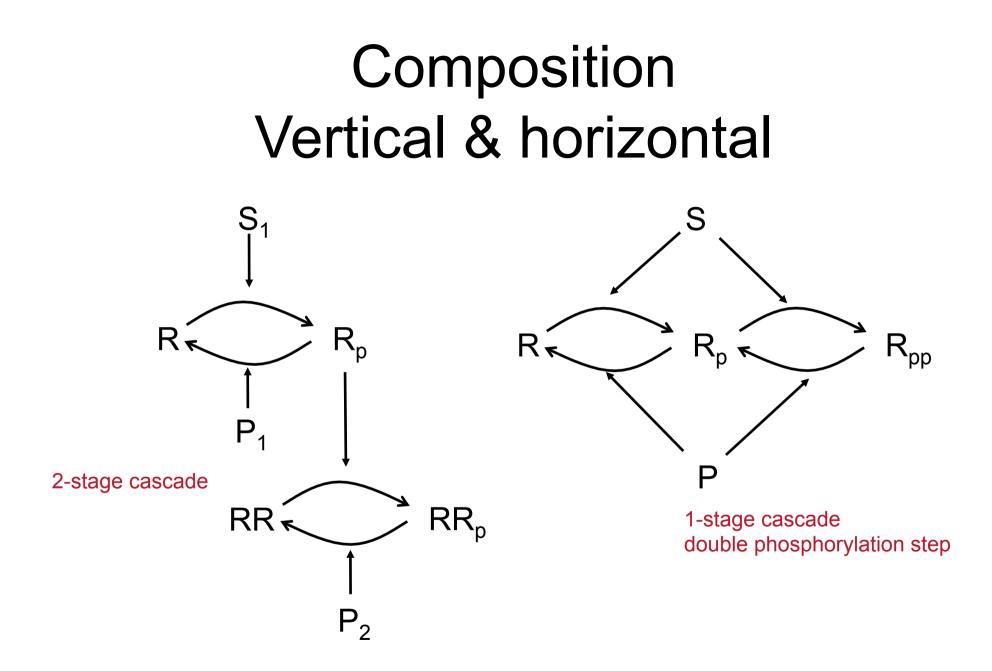
R

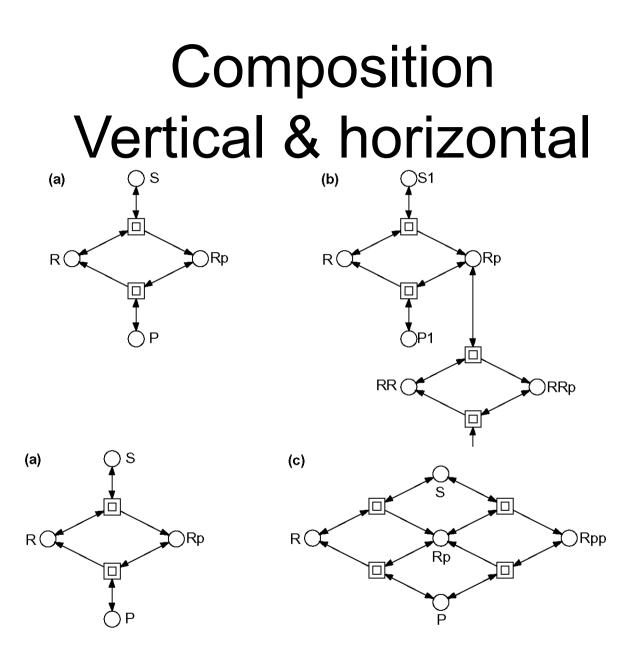
Questions

- Is Michaelis-Menten adequate for phosphorylation pathways?
- Is Mass Action sufficient/correct for these pathways?
- What is the effect of negative feedback?
- Can we confirm the 'negative feedback amplifer' behaviour in both MM and MA models
- Can oscillators be built?
- Overall, what are the rules for component-based construction?

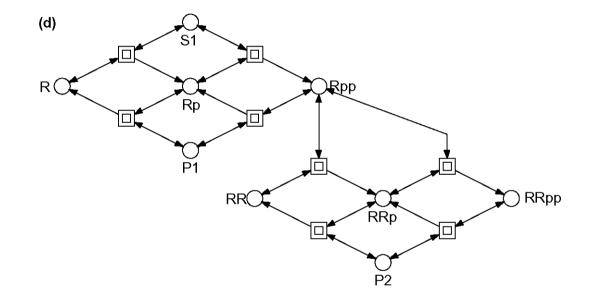


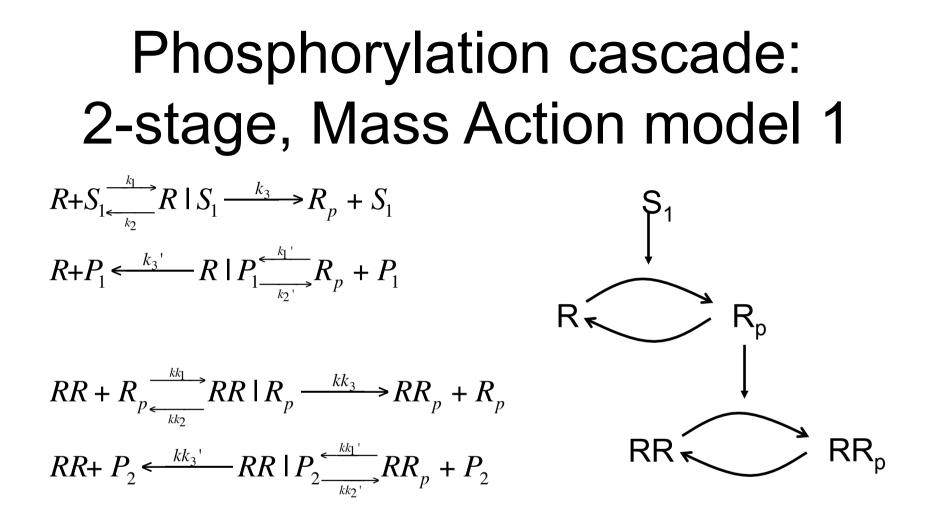
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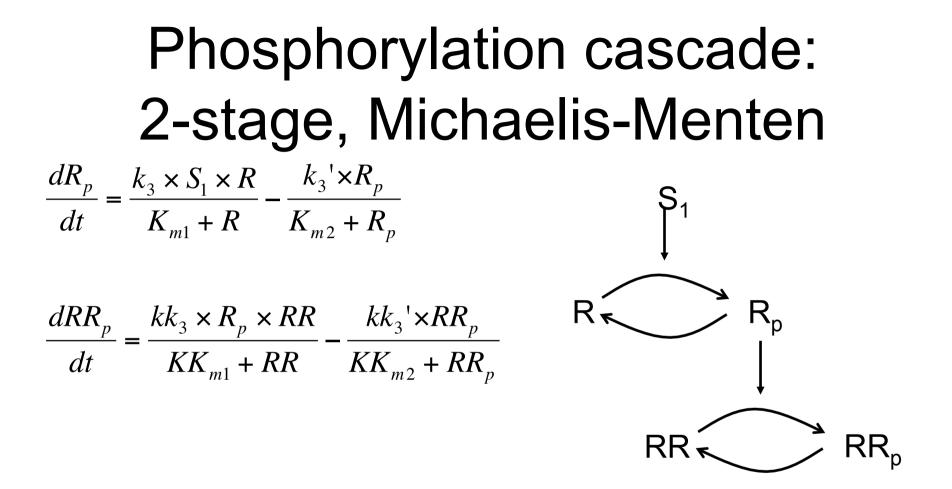




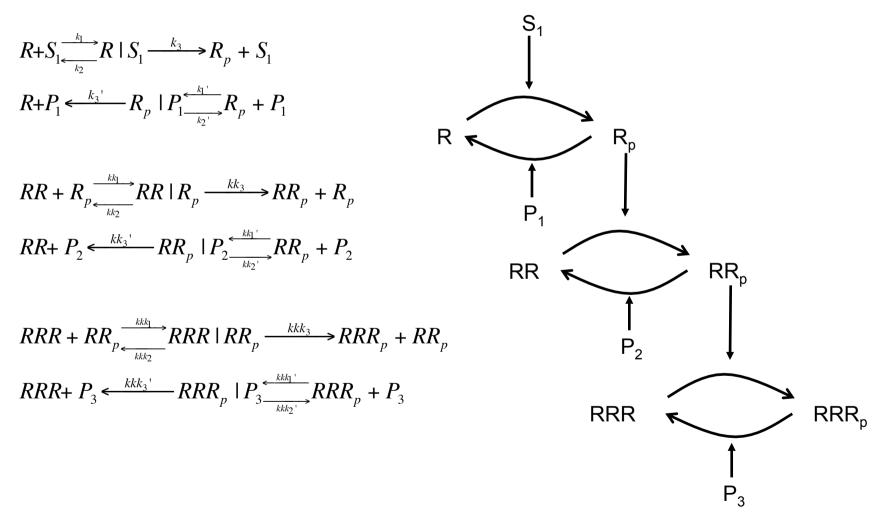
Two stage, double phosphorylation







3-stage Phosphorylation cascade (Mass Action)



Engineering Biochemical Network models

Phosphorylation cascade: 3-stage, Michaelis-Menten

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$$\frac{dR_{p}}{dt} = \frac{k_{3} \times S_{1} \times R}{K_{m1} + R} - \frac{k_{3}' \times R_{p}}{K_{m2} + R_{p}}$$

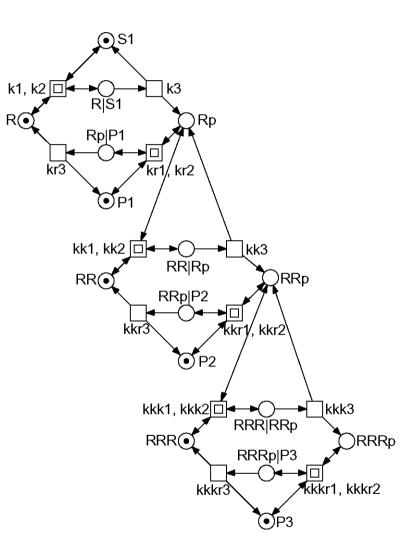
$$\frac{dRR_{p}}{dt} = \frac{kk_{3} \times R_{p} \times RR}{KK_{m1} + RR} - \frac{kk_{3}' \times RR_{p}}{KK_{m2} + RR_{p}}$$

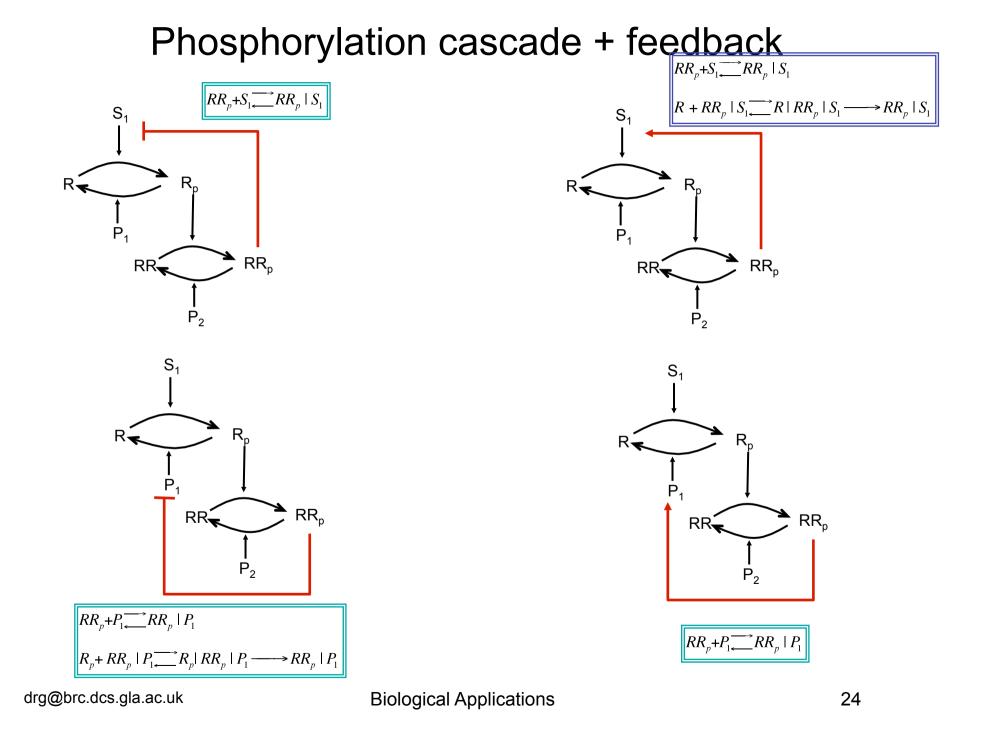
$$\frac{dRRR_{p}}{dt} = \frac{kkk_{3} \times RR_{p} \times RRR}{KKK_{m1} + RRR} - \frac{kkk_{3}' \times RRR_{p}}{KKK_{m2} + RRR_{p}}$$

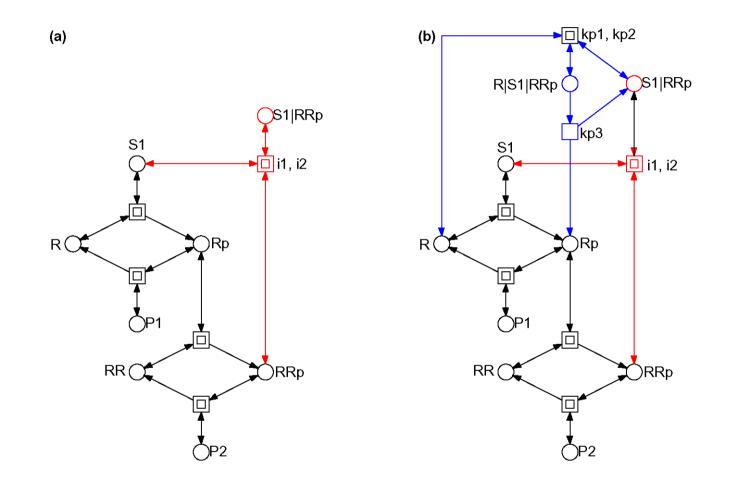
$$RR \longrightarrow RRR_{p}$$

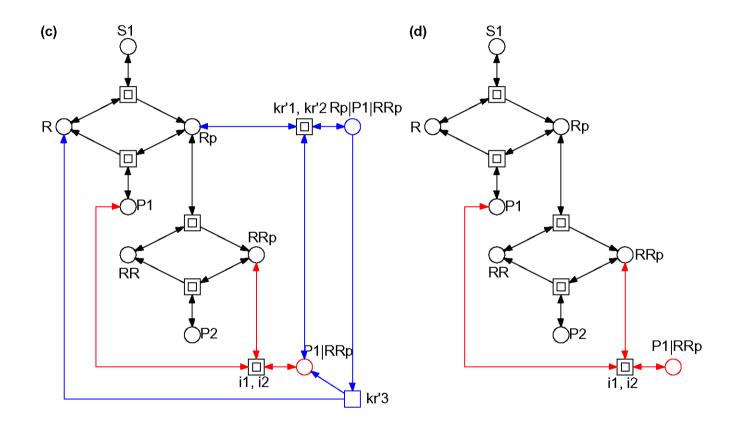
$$RRR \longrightarrow RRR_{p}$$

3-stage

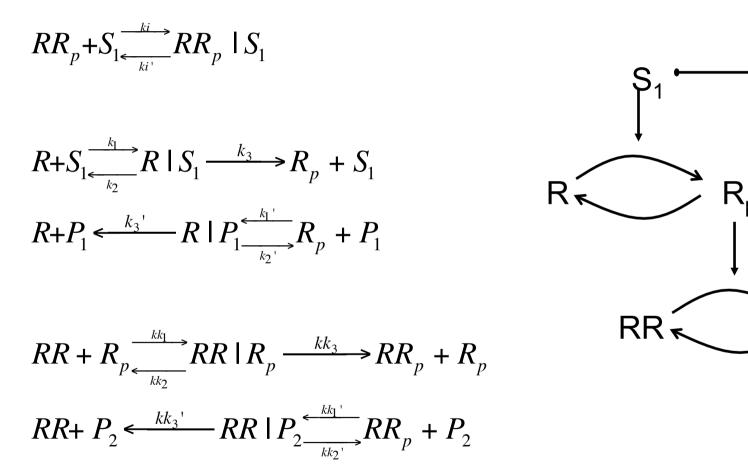








Phosphorylation cascade + negative feedback: 2-stage, Mass Action model 1



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RR_n

Phosphorylation cascade + negative feedback: 2-stage, Michaelis-Menten

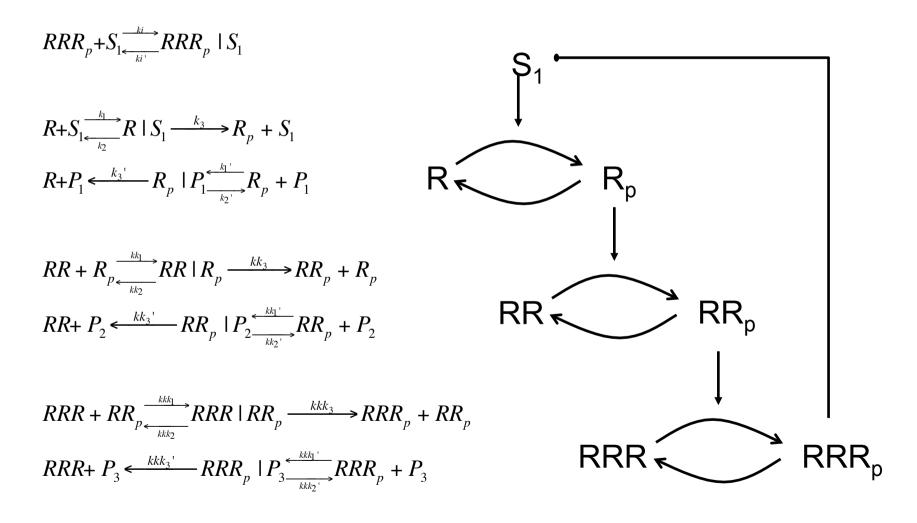
$$\frac{dR_{p}}{dt} = \frac{k_{3} \times S_{1} \times R}{K_{m1} \times \left(1 + \frac{RR_{p}}{K_{i}}\right) + R} - \frac{k_{3}' \times R_{p}}{K_{m2} + R_{p}}$$

$$\frac{dRR_{p}}{dt} = \frac{kk_{3} \times R_{p} \times RR}{KK_{m1} + RR} - \frac{kk_{3}' \times RR_{p}}{KK_{m2} + RR_{p}}$$

$$R = \frac{kk_{3} \times R_{p} \times RR}{RR} = \frac{kk_{3} \times R_{p} \times RR}{RR} = \frac{kk_{3}' \times RR_{p}}{RR}$$

- Using Competitive Inhibition
- $V = V_{\max} \times \frac{[S]}{[S] + K_m \times \left(1 + \frac{[I]}{[K_i]}\right)}$ Ki is the dissociation constant for the SI complex •

Phosphorylation cascade + negative feedback: 3-stage, Mass Action, model 1



Phosphorylation cascade + negative feedback: 3-stage, Michaelis-Menten

S.

$$\frac{dR_{p}}{dt} = \frac{k_{3} \times S_{1} \times R}{K_{m1} \times \left(1 + \frac{RRR_{p}}{K_{i}}\right) + R}} - \frac{k_{3}' \times R_{p}}{K_{m2} + R_{p}}$$

$$\frac{dRR_{p}}{dt} = \frac{kk_{3} \times R_{p} \times RR}{KK_{m1} + RR}} - \frac{kk_{3}' \times RR_{p}}{KK_{m2} + RR_{p}}$$

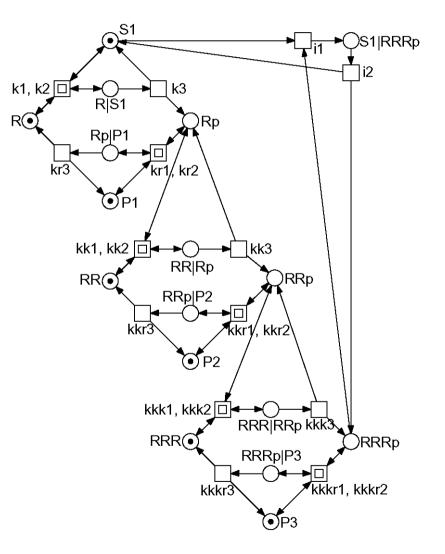
$$\frac{dRRR_{p}}{dt} = \frac{kkk_{3} \times RR_{p} \times RRR}{KKK_{m1} + RRR}} - \frac{kkk_{3}' \times RRR_{p}}{KKK_{m2} + RRR_{p}}$$

$$\frac{kk_{3} \times RR_{p} \times RRR}{KKK_{m1} + RRR}} - \frac{kkk_{3}' \times RRR_{p}}{KKK_{m2} + RRR_{p}}$$

$$\frac{V = V_{max}}{S} \times \frac{[S]}{[S] + K_{m} \times \left(1 + \frac{[I]}{[K_{i}]}\right)}$$

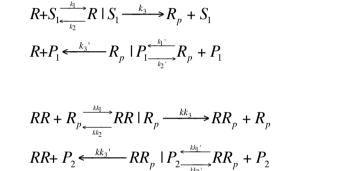
- Using Competitive Inhibition ۲
- Ki is the dissociation constant for the SI complex •

3-stage, negative feedback



Phosphorylation cascade + negative feedback: 3-stage, Inhibitor on 2nd stage, Mass Action

 $RRR_{p} + S_{1 \leftarrow ki'} RRR_{p} \mid S_{1}$



$$RRR + RR_{p} \xrightarrow{kkk_{1}} RRR \mid RR_{p} \xrightarrow{kkk_{3}} RRR_{p} + RR_{p}$$
$$RRR + P_{3} \xleftarrow{kkk_{3}} RRR_{p} \mid P_{3} \xrightarrow{kkk_{1}} RRR_{p} + P_{3}$$

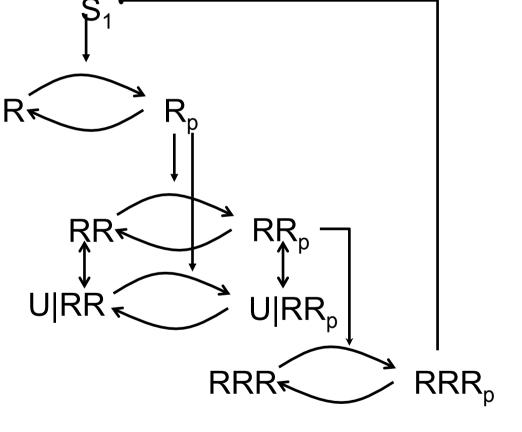
$$U + RR \xrightarrow[ku_{1}]{ku_{2}} U | RR$$

$$U + RR_{p} \xrightarrow[ku_{2}]{ku_{2}} U | RR_{p}$$

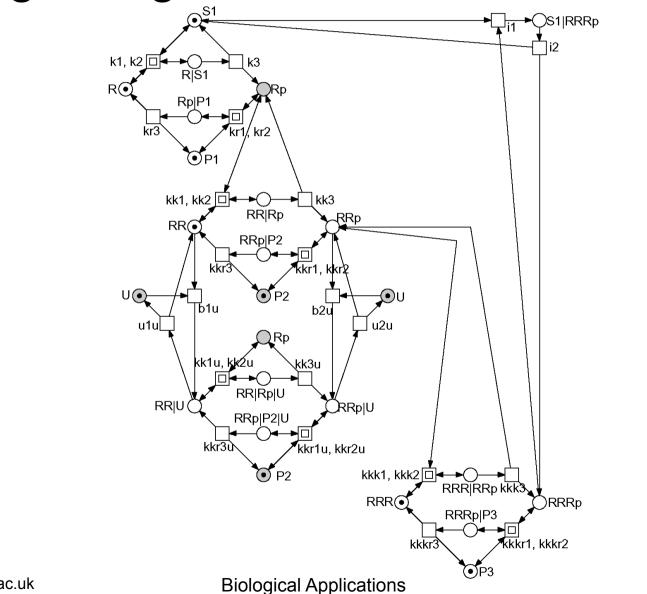
$$U | RR + R_{p} \xrightarrow[ku_{2}]{ku_{2}} U | RR | R_{p} \xrightarrow{kk_{3}} U | RR_{p} + R_{p}$$

$$U | RR + P_{2} \xleftarrow{kk_{3}'} U | RR | R_{p} | P_{2} \xrightarrow[kk_{2}]{kk_{2}} U | RR_{p} + P_{2}$$

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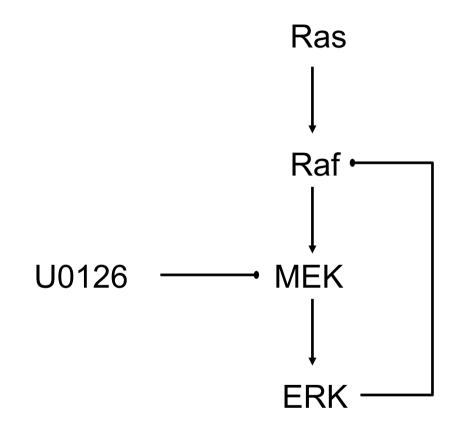


3-stage, negative feedback + inhibitor



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'Real cascade & feedback'

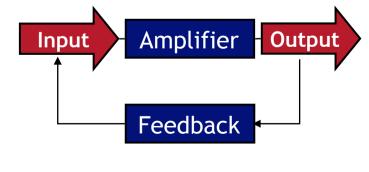


Is the ERK pathway a negative feedback amplifier?

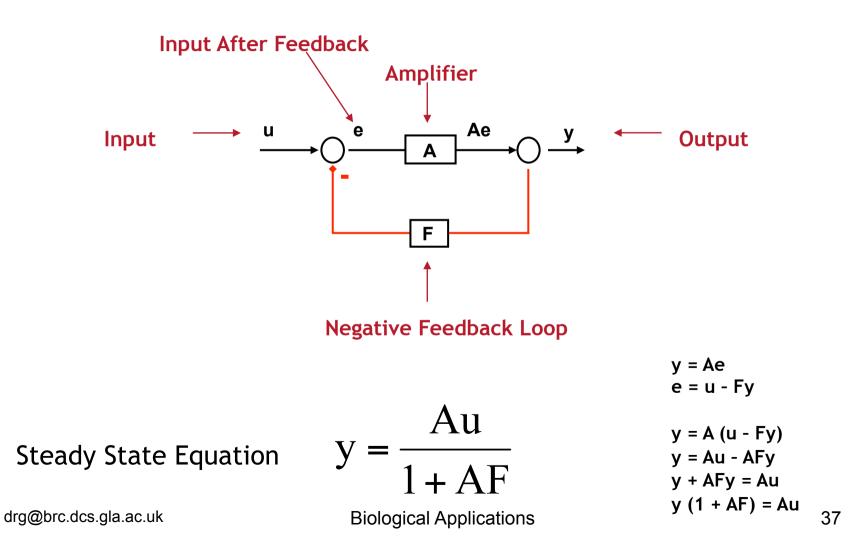
Sauro HM, Kholodenko BN. Quantitative analysis of signaling networks. Prog Biophys Mol Biol. 2004 Sep;86(1):5-43.

Negative Feedback Amplifier

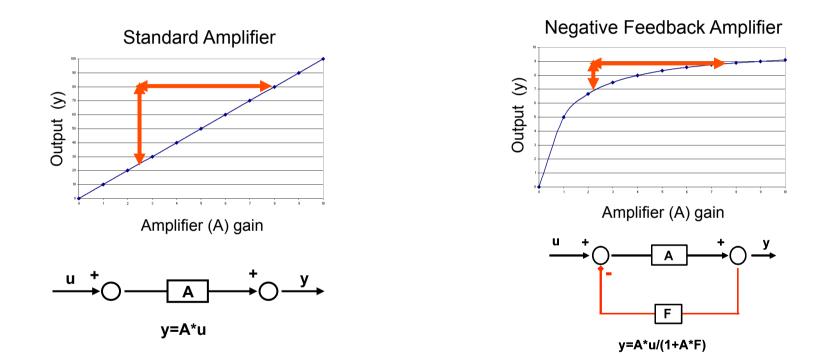
- A negative feedback amplifier stems from the field of electronics and consists of an amplifier with a negative feedback loop from the output of the amplifier to its input.
- The negative feedback loop results in a system that is much more robust to disturbances in the amplifier.
- The negative feedback amplifier was invented in 1927 by Harold Black of Western Electric and was originally used for reducing distortion in long distance telephone lines.
- The negative feedback amplifier is now a key electrical component used in a wide variety of applications



Negative Feedback Amplifier

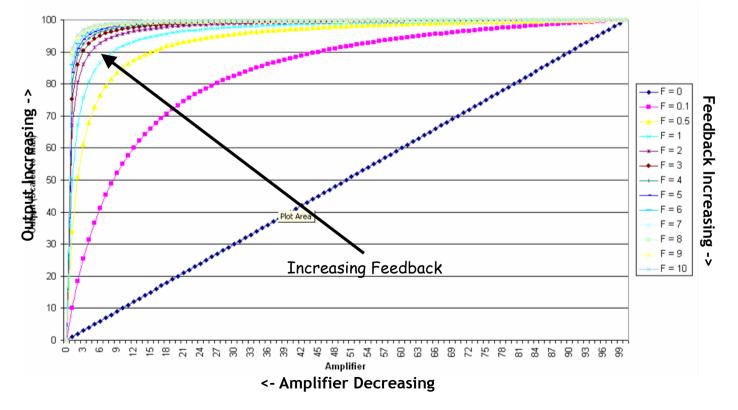


The negative feedback imparts signalling robustness



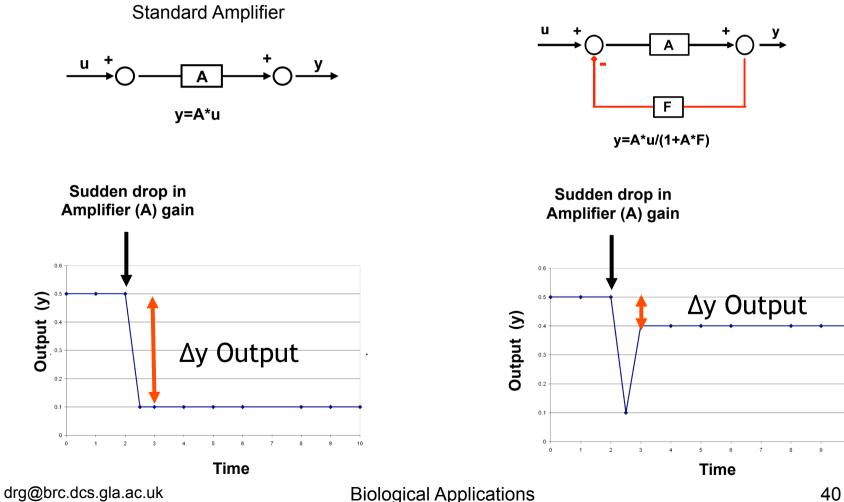
A large change in amplifier gain leads to a small change in output (y)

Feedback



The negative feedback imparts signalling robustness

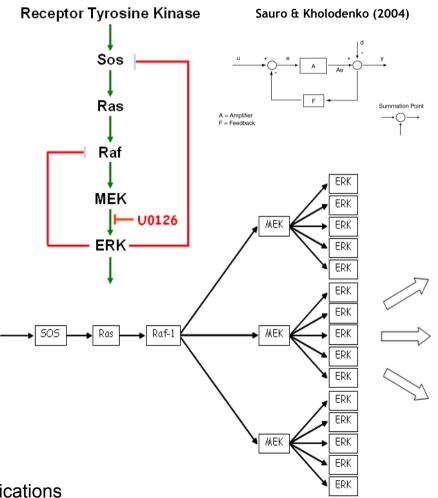
Negative Feedback Amplifier



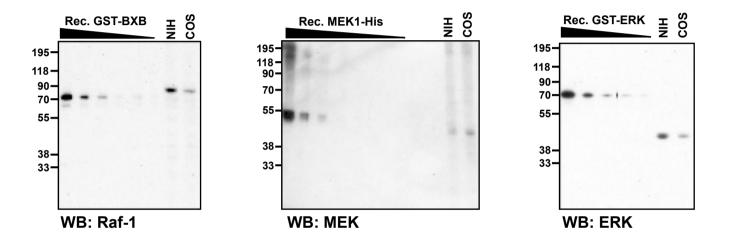
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Application to Biology

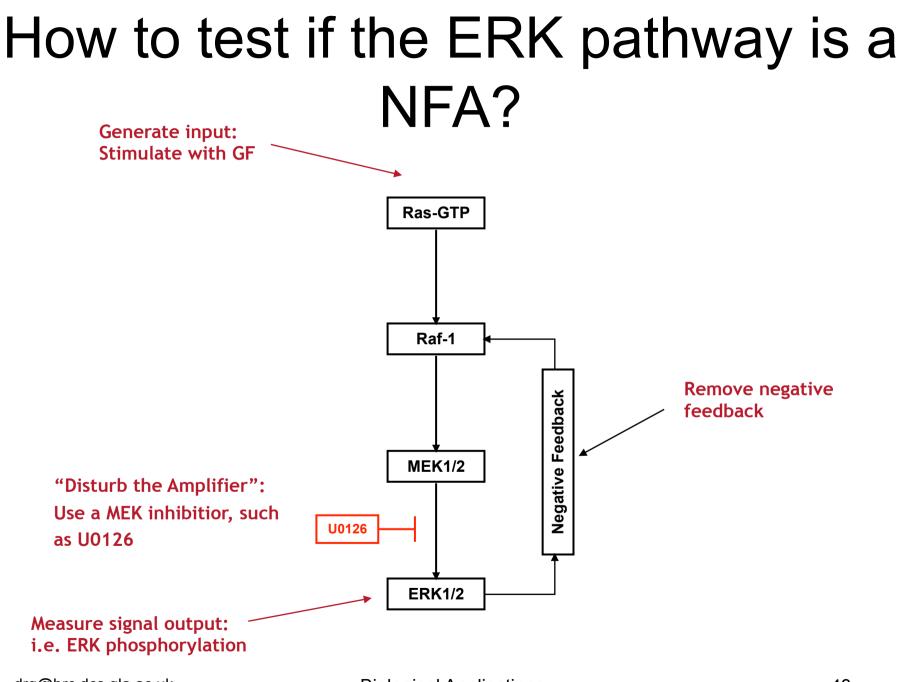
- The ERK cascade is a well known biological amplifier and contains numerous negative feedback loops.
- At first sight, it has the correct structure to be a negative feedback amplifier.
- If the ERK cascade is a negative feedback amplifier it should be robust to disturbances within the cascade.
- From a biological point of view, these disturbances could be caused by drugs, such as U0126, aimed at decreasing the activity of the ERK cascade.
- This suggests that these drugs will be relatively ineffective.
- In fact, current drugs aimed at decreasing the activity of the MAPK pathway have proved less efficient in *in vivo* applications than anticipated from *in vitro* inhibition assays.



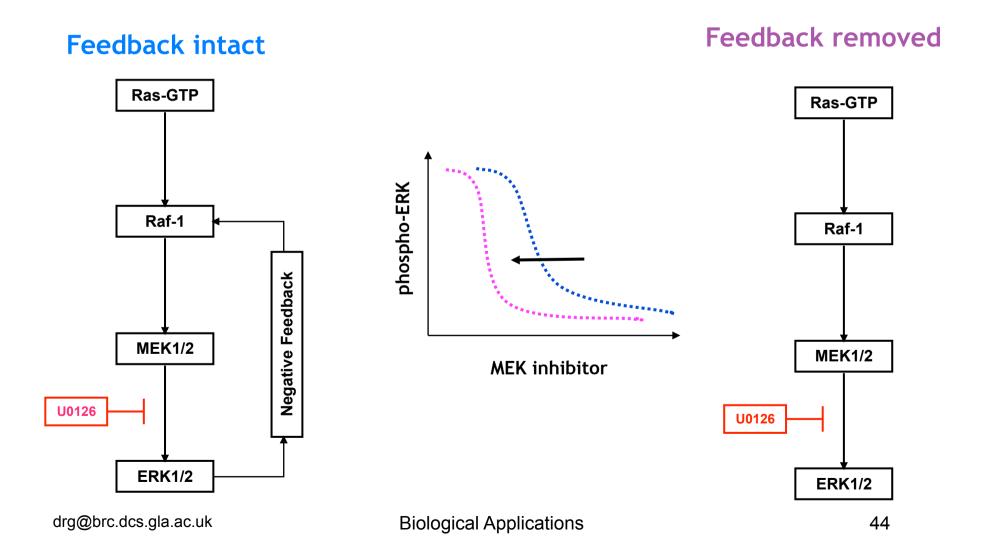
Raf/MEK/ERK amplifies the signal



Cell line	Raf-1	MEK	ERK	Concentration per cell
COS1	3.6	10.6	21.2	femtomol
	1	2.9	5.9	ratio
NIH 3T3	10.9	7.1	98	femtomol
	1	0.7	9	ratio



Hypothesis: Braking the feedback should sensitise the ERK pathway to MEK-inhibitor



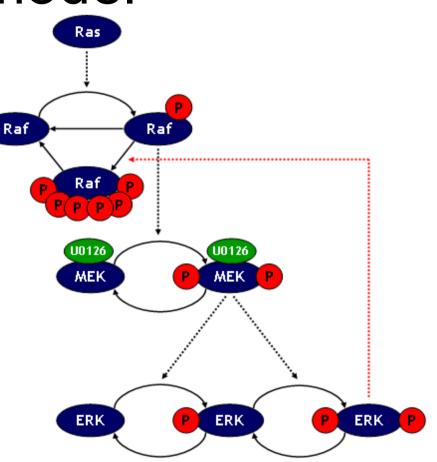
How to test if the ERK pathway is a NFA?

Strategy

In vivo system that allows us to compare feedback broken to feedback intact model. Computational Model of ERK pathway with/without feedback

Computational Modeling 1: Build the model

- Non-linear ordinary differential equations (ODE's).
- ODE's were solved using Math Lab and Gepasi.
- Models are based on the Schoeberl et al. (2002) model
- Mass Action Kinetics instead of Michaelis Menten
- Kinetic parameters are from literature, previous models and "guesstimates"



Schoeberl et al. (2002), Computational modeling of the dynamics of the MAP kinase cascade activated
by surface and internalized EGF receptors, Nature Biotechnology 20, 370-375drg@brc.dcs.gla.ac.ukBiological Applications46

Amplifier / negative feedback

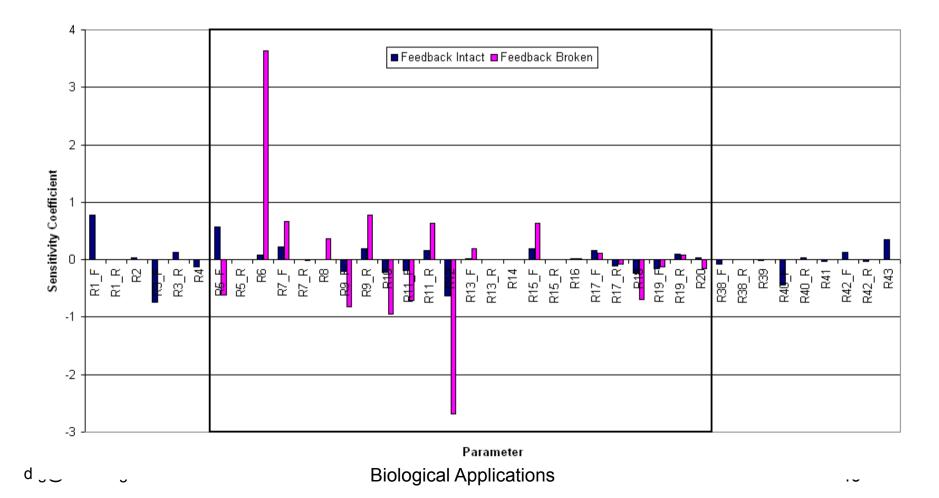
- Model amplifier strength by
 - Adding inhibitor to 2nd stage
 - Modifying kk3, kkk3 [I.e. modifying rate of production of RRp, RRRp]
 - Add/remove cascade elements
- Then plot amp strength versus output, e.g. [U] vs [RRRp]
- ?Model feedback strength by
 - Leaving out feedback loop
 - varying ki, and plot ki vs [RRRp]
- Notes: avoid saturation; use signal in linear range; ? model degradation in S1 signal?

Computational Modeling 2: Results Feedback intact Feedback broken RafP RafBXB-EF Phase4 RasGTP Phase1 Rafx MEK MEK-P MEK-PP MEK MEK-P MEK-PP Phase2 Phase2 13000000 ERK ERK-P ERK-PP ERK ERK-P ERK-PP 11000000 Total Steady State [ERK-PP] Phase3 Phase3 9000000 7000000 5000000 Plot Area 3000000 1000000 5000000 10000000 15000000 20000000 -1000000 Initial [U0126]

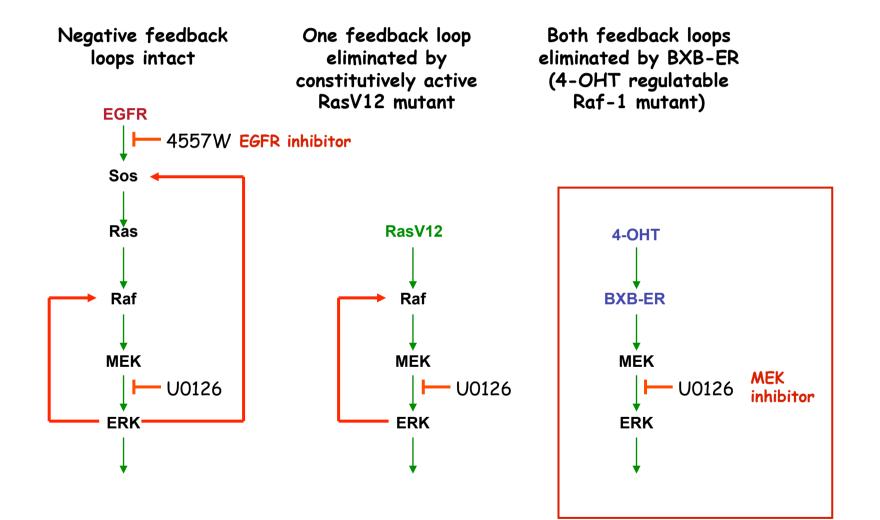
Prediction: Braking the feedback modulates drug response drg@brc.dcs.gla.ac.uk Biological Applications 48

Computational Modeling 2:

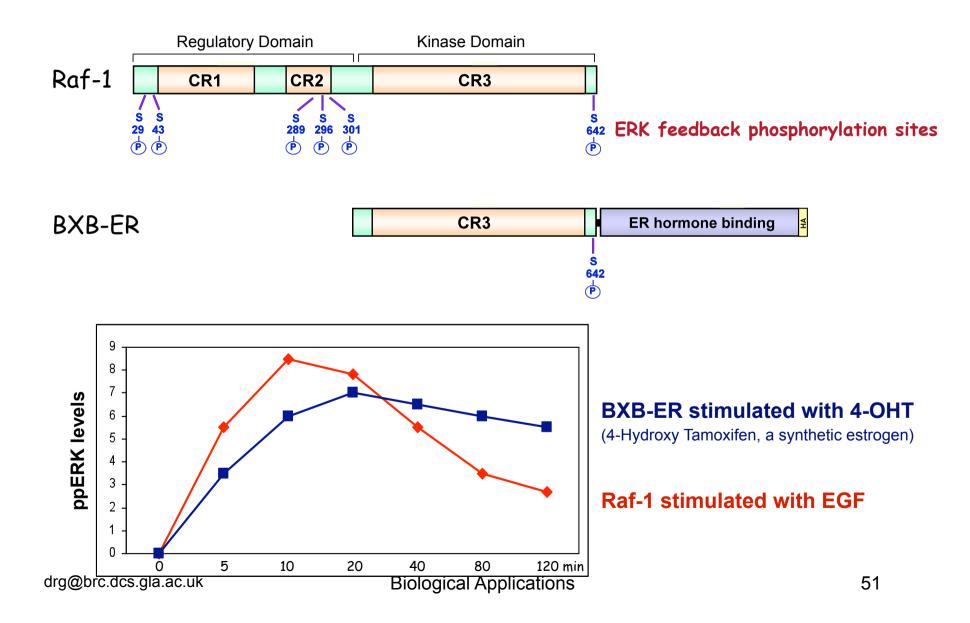
Sensitivity of kine Restantesers is decreased due to Negative Feedback



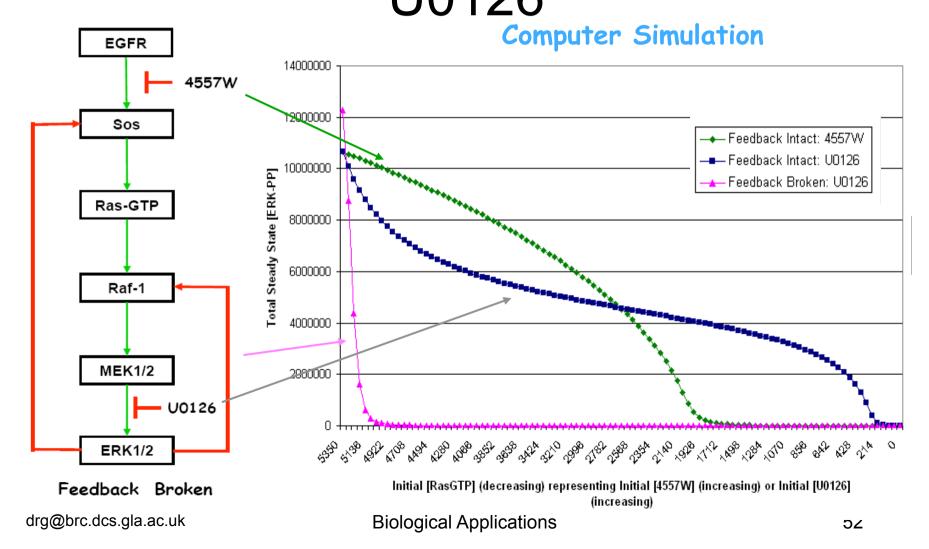
The experimental systems



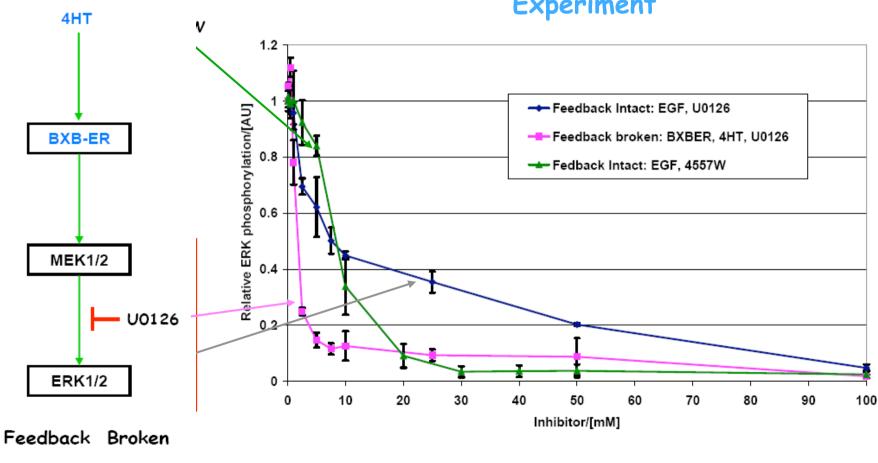
Breaking the ERK feedback with BXBER



Ablation of feedback by BXBER decreases robustness to MEK-inhibitor U0126



Ablation of feedback by BXBER decreases robustness to MEKinhibitor U0126

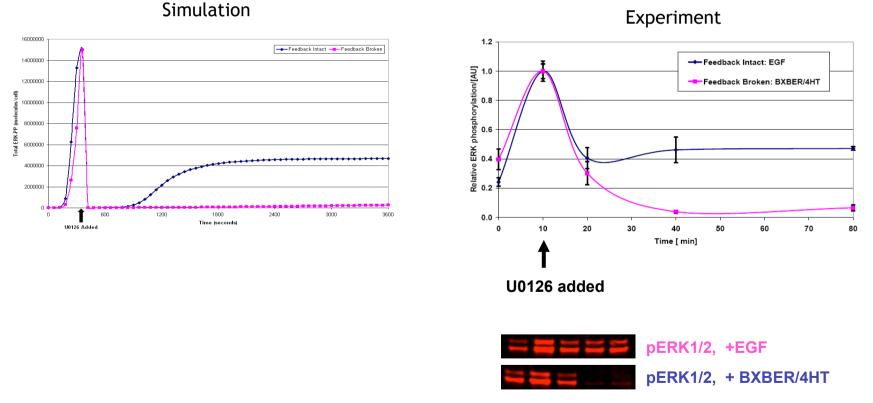


Experiment

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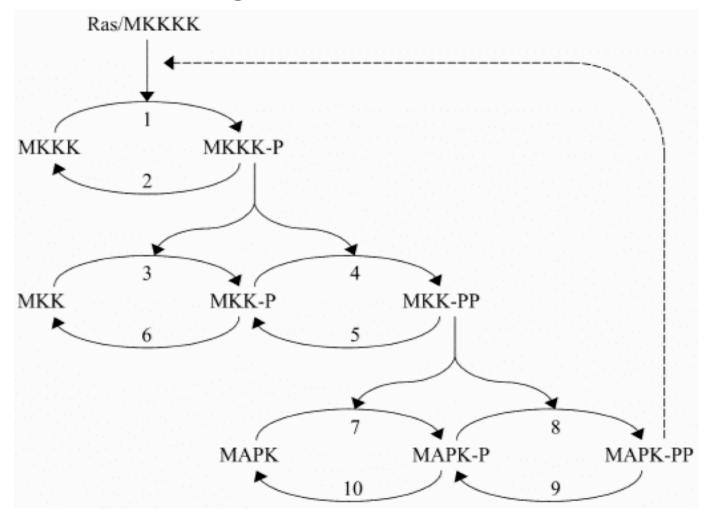
Signal recovery after MEK inhibition



0 10 20 40 80 min stimulation

Biological Applications

Kholodenko – negative feedback oscillator

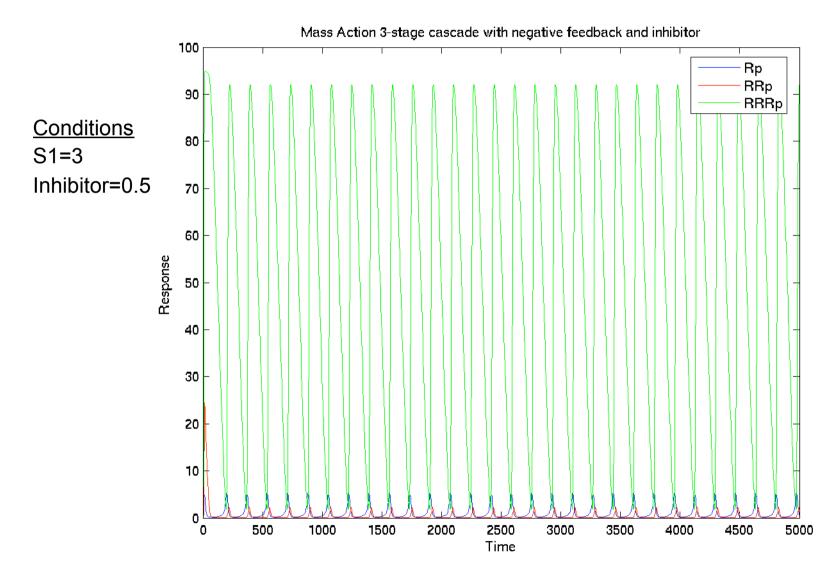


Negative feedback and ultrasensitivity can bring about oscillations in the mitogen-activated protein kinase cascades. Kholodenko BN., Eur J Biochem 2000 Mar;267(6):1583-8

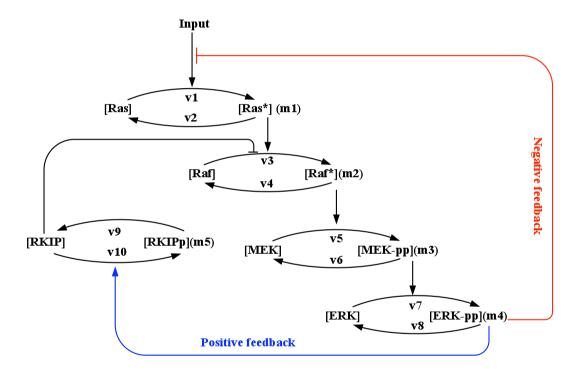
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Oscillations! Phosphorylation cascade + negative feedback: 3-stage, Inhibitor on 2nd stage, Mass Action

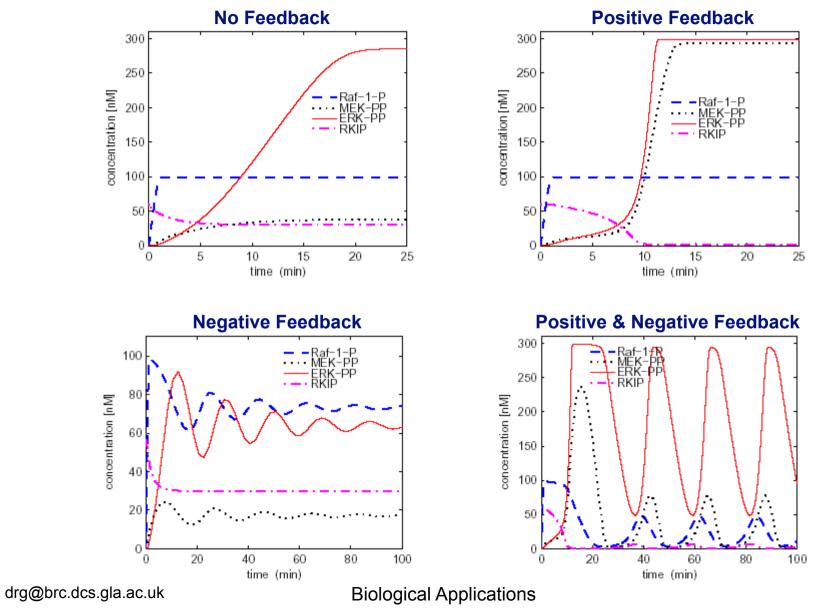


Combination of positive & negative feedback Mathematical Model



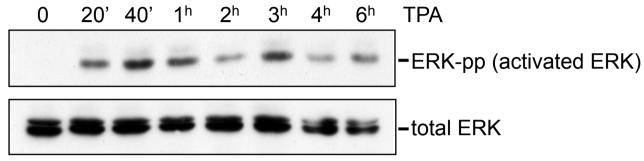
Modeling and Analysis of Two Feedback Loop Dynamics in Ras/Raf-1/MEK/ERK Signaling Pathway Kwang-Hyun Cho, Sung-Young Shin, Walter Kolch, Olaf Wolkenhauer. ICSB 2004

Combination of positive & negative feedback: Simulation



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Combination of positive & negative feedback: Simulation vs. Experimental Data



Western blots COS1 cell lysates

